

***** STN Columbus *****

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=> index bioscience medicine

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=> S (hydroxynitrile (w) lyase) or hydroxynitrilase)

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46 FILE WPINDE
1 FILE NLDB

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L1 QUE ((HYDROXYNITRILE (W) LYASE) OR HYDROXYNITRILASE)

=> d rank

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F41	1	WPIFV
F42	1	NLDB

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FILE 'WPIDS' ENTERED AT 12:02:19 ON 17 NOV 2008
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=> S L1
L2 1409 L1

=> S (modif? or mutant or variant or substitut?)(s) L2
L3 157 (MODIF? OR MUTANT OR VARIANT OR SUBSTITUT?)(S) L2

=> S cyanohydrin (s) L3
L4 44 CYANOHYDRIN (S) L3

=> S (cassava or manihot) (s) L4
L5 23 (CASSAVA OR MANIHOT) (S) L4

=> S (cassava or manihot) and L4
L6 29 (CASSAVA OR MANIHOT) AND L4

=> dup rem L6
PROCESSING COMPLETED FOR L6
L7 13 DUP REM L6 (16 DUPLICATES REMOVED)

=> D ibib abs L7 1-13

L7 ANSWER 1 OF 13 USPATFULL on STN
ACCESSION NUMBER: 2008:143550 USPATFULL <<LOGINID::20081117>>
TITLE: Novel Modified S-Hydroxynitrile Lyase
INVENTOR(S): Ichige, Eita, Chiba, JAPAN
Semba, Hisashi, Ibaraki, JAPAN
Shijuku, Toshiaki, Chiba, JAPAN
Harayama, Shigeaki, Chiba, JAPAN

NUMBER	KIND	DATE

PATENT INFORMATION:	US 20080124784	A1 20080529
APPLICATION INFO.:	US 2005-594732	A1 20050330 (10)
	WO 2005-JP6730	20050330
		20060929 PCT 371 date

NUMBER	DATE

PRIORITY INFORMATION:	JP 2004-105642 20040331
DOCUMENT TYPE:	Utility
FILE SEGMENT:	APPLICATION
LEGAL REPRESENTATIVE:	FENNIGAN, HENDERSON, FARABOW, GARRETT & DUNNER, LLP, 901 NEW YORK AVENUE, NW, WASHINGTON, DC, 20001-4413, US
NUMBER OF CLAIMS:	12
EXEMPLARY CLAIM:	1
NUMBER OF DRAWINGS:	21 Drawing Page(s)
LINE COUNT:	3024
CAS INDEXING IS AVAILABLE FOR THIS PATENT.	
AB	This invention relates to S-hydroxynitrile lyase having excellent tolerance to heat, organic solvents, and the like, which is obtained by

modifying at least one amino acid in the helix D3', helix A, and
beta-sheet 2 domains in the amino acid sequence of wild-type
S-hydroxynitrile lyase.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L7 ANSWER 2 OF 13 Elsevier BIOBASE COPYRIGHT 2008 Elsevier Science B.V. on
STN DUPLICATE

ACCESSION NUMBER: 2008085250 ESI/BIOS <<LOGINID::20081117>>

TITLE: Hydroxynitrile lyase catalyzed cyanohydrin synthesis
at high pH-values

AUTHOR: Von Langermann J.; Guterl J.-K.; Pohl M.; Wajant H.;
Kragl U.

CORPORATE SOURCE: U. Kragl, Institut für Chemie, Universität Rostock,
Albert-Einstein-Str. 3a, 18059 Rostock, Germany.
E-mail: udo.kragl@uni-rostock.de

SOURCE: Bioprocess and Biosystems Engineering. (2008), 31/3
(155-161), 28 reference(s)
CODEN: BBEBV ISSN: 1615-7591

DOCUMENT TYPE: Journal, Article

COUNTRY: Germany, Federal Republic of

LANGUAGE: English

SUMMARY LANGUAGE: English

AB The application of unusual high pH-values within enzymatic
cyanohydrin synthesis has been investigated. Usually enzymatic
cyanohydrin synthesis in two-phase systems requires low pH-values
within the aqueous phase to suppress the non-enzymatic side reaction. In
contrast, we investigated the usage of pH-values above pH 6 by using the
highly enantioselective (S)-selective ***hydroxynitrile***
lyase from ***Manihot*** esculenta. With these unusual
reaction conditions also the unfavorable substrate 3-phenoxy-benzaldehyde
can be converted by the wild type enzyme with excellent conversion and
enantiomeric excess yielding pure (S)-3-phenoxy-benzaldehyde
cyanohydrin with an enantiomeric excess of 97%. Although the
variant MeHNL-W128A shows a higher activity with respect to this
reaction, the enantioselectivity was reduced (85% e.e.(S)). Additionally,
a new continuous spectroscopic ***cyanohydrin*** assay monitoring the
formation of 3-phenoxy-benzaldehyde ***cyanohydrin*** was developed.
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L7 ANSWER 3 OF 13 WPI/DS COPYRIGHT 2008 THOMSON REUTERS on STN

ACCESSION NUMBER: 2007-361592 [34] WPI/DS

DOC. NO. CPI: C2007-131327 [34]

TITLE: Novel ***modified*** S- ***hydroxynitrile***
lyase SHNL having acid resistance and obtained by
substituting amino acids in wild-type SHNL
sequence derived from ***cassava*** or para rubber
tree, useful for producing optically active
cyanohydrin

DERWENT CLASS: B05; D16; E16

INVENTOR: HARAYAMA S; ICHIGE E; SEMBA H; SHIUKU T; SENBA T; YOSOKU
T

PATENT ASSIGNEE: (JAPC-C) NIPPON SHOKUBAI CO LTD; (IDKU-N) DOKURITSU
GYOSEI HOJIN SEIHIN HYOKA GIJU; (NATE-N) NAT INST
TECHNOLOGY & EVALUATION

COUNTRY COUNT: 116

PATENT INFO ABBR.:

PATENT NO KIND DATE WEEK LA PG MAIN IPC

WO 2007037354 A1 20070405 (200734) JA 65[26]

JP 2007089513 A 20070412 (200734) JA 33

EP 1944366 A1 20080716 (200849) EN

APPLICATION DETAILS:

PATENT NO KIND APPLICATION DATE

WO 2007037354 A1 WO 2006-JP319422 20060929

JP 2007089513 A	JP 2005-285049 20050929
EP 1944366 A1	EP 2006-810828 20060929
EP 1944366 A1	WO 2006-JP319422 20060929

FILING DETAILS:

PATENT NO	KIND	PATENT NO
EP 1944366	A1 Based on	WO 2007037354 A

PRIORITY APPLN. INFO: JP 2005-285049 20050929

AN 2007-361592 [34] WPIDS

AB WO 2007037354 A1 UPAB: 20070529

NOVELTY - A modified S-hydroxynitrile lyase (SHNL) obtained by substituting at least one amino acid chosen from amino acid at position 36, 140 and 209 in amino acid sequence (SEQ ID No. 2) of wild-type SHNL derived from *Manihot* (*Manihot* esculenta), or substituting at least one amino acid chosen from amino acid at position 36, 139 and 208 in amino acid sequence (SEQ ID No. 3) of wild-type SHNL derived from para rubber tree (*Hevea brasiliensis*), is new.

DETAILED DESCRIPTION - A modified S-hydroxynitrile lyase (SHNL) obtained by substituting at least one amino acid chosen from amino acid at position 36, 140 and 209 in amino acid sequence (SEQ ID No. 2) of wild-type SHNL derived from *Manihot* (*Manihot* esculenta), where leucine at position 36 is substituted to methionine, threonine at position 140 to isoleucine, and lysine at position 209 to asparagine; or substituting at least one amino acid chosen from amino acid at position 36, 139 and 208 in amino acid sequence (SEQ ID No. 3) of wild-type SHNL derived from para rubber tree (*Hevea brasiliensis*).

INDEPENDENT CLAIMS are included for the following:

- (1) DNA encoding the modified SHNL; and
- (2) method for producing the modified SHNL.

USE - For producing optically active cyanohydrin, which involves contacting the modified SHNL with carbonyl compound and cyanide compound (claimed).

ADVANTAGE - The modified SHNL has improved acid resistance compared to wild-type SHNL, and enables efficient production of optically active cyanohydrin by suppressing racemization reaction in acidic conditions.

L7 ANSWER 4 OF 13 WPIDS COPYRIGHT 2008 THOMSON REUTERS on STN

ACCESSION NUMBER: 2006-294347 [30] WPIDS

DOC. NO. CFI: C2006-096253 [30]

TITLE: *Manihot* mutant *Manihot* hydroxynitrile *Manihot* lyase

, useful for efficiently producing large quantities of *Manihot* cyanohydrin and hydroxycarboxylic acid

DERWENT CLASS: B05; D16

INVENTOR: AKIYAMA T; ASANO Y; SATO E; YU F

PATENT ASSIGNEE: (MITR-C) MITSUBISHI RAYON CO LTD

COUNTRY COUNT: 111

PATENT INFO ABBR.:

PATENT NO	KIND	DATE	WEEK	LA	PG	MAIN IPC
WO 2006041226	A1	20060420 (200630)*	JA	161	[11]	
EP 1811028	A1	20070725 (200750)	EN			
JP 2006541007	X	20080522 (200836)	JA	79		

APPLICATION DETAILS:

PATENT NO	KIND	APPLICATION	DATE
WO 2006041226	A1	WO 2005-JP19360	20051014
EP 1811028	A1	EP 2005-795304	20051014
EP 1811028	A1	WO 2005-JP19360	20051014
JP 2006541007	X	WO 2005-JP19360	20051014
JP 2006541007	X	JP 2006-541007	20051014

FILING DETAILS:

PATENT NO	KIND	PATENT NO
EP 1811028	A1 Based on	WO 2006041226 A
JP 2006541007	X Based on	WO 2006041226 A

PRIORITY APPLN. INFO: JP 2005-58857 20050303

JP 2004-301718 20041015

JP 2004-355766 20041208

AN 2006-294347 [30] WPI/D5

AB WO 2006041226 A1 UPAB: 20060510

NOVELTY - A mutant hydroxynitrile lyase (I) comprising at least one amino acid substitution, is new. The amino acid substitution is preferably replacement of histidine 103 of the wild-type sequence by another amino acid and/or replacement of at least one lysine of the wild-type sequence by another amino acid.

DETAILED DESCRIPTION - An improved hydroxynitrile lyase (I) chosen from (a) improved hydroxynitrile lyase obtained by substituting amino acid residue at second position in the wild-type hydroxynitrile lyase with another amino acid, (b) improved hydroxynitrile lyase obtained by substituting amino acid residue at position 103 or histidine at position near 103 in the wild-type hydroxynitrile lyase with another amino acid, (c) improved hydroxynitrile lyase obtained by substituting at least one lysine residue in the wild-type hydroxynitrile lyase with another amino acid, (d) improved hydroxynitrile lyase obtained by substituting amino acid residue at second position and amino acid residue at position 103 or histidine at position near 103 in the wild-type hydroxynitrile lyase with another amino acid, (e) improved hydroxynitrile lyase obtained by substituting amino acid residue at second position and at least one lysine residue in the wild-type hydroxynitrile lyase with another amino acid, (f) improved hydroxynitrile lyase obtained by substituting at least one lysine residue and amino acid residue at position 103 or histidine at position near 103 in the wild-type hydroxynitrile lyase with another amino acid, and (g) improved hydroxynitrile lyase obtained by substituting at least one lysine residue, amino acid residue at second position, and amino acid residue at position 103 or histidine at position near 103 in the wild-type hydroxynitrile lyase with another amino acid.

INDEPENDENT CLAIMS are also included for the following:

- (1) improved type hydroxynitrile lyase gene (II) encoding (I);
- (2) recombinant vector (III) containing (II);
- (3) transformant (IV) obtained by introducing (III) into a host;
- (4) culture (V) obtained by cultivating (IV);
- (5) producing (M1) cyanohydrin, involves processing ketone or aldehyde compound and a cyanide compound with (I), and extracting cyanohydrin from the processed substance; and
- (6) producing hydroxycarboxylic acid, involves hydrolyzing cyanohydrin obtained by (M1).

USE - (I) or (M1) is useful for producing cyanohydrin (claimed), which is useful for synthesizing various optically active intermediates used for the synthesis of alpha hydroxy acid, alpha hydroxy ketone, and beta amino alcohol, and useful in pharmaceuticals. (I) is useful for producing hydroxy carboxylic acid.

ADVANTAGE - (I) has improved hydroxynitrile lyase activity, can be efficiently produced in large quantities, and enables effective production of cyanohydrin.

L7 ANSWER 5 OF 13 USPATFULL on STN

ACCESSION NUMBER: 2005:105041 USPATFULL <<LOGINID::20081117>>

TITLE: Enzyme reaction method and a method for enzymatically

producing an optically active cyanohydrin

INVENTOR(S): Semba, Hisashi, Ibaraki, JAPAN

Dobashi, Yukio, Ibaraki, JAPAN

PATENT ASSIGNEE(S): NIPPON SHOKUBAI CO., LTD. (non-U.S. corporation)

NUMBER	KIND	DATE

PATENT INFORMATION: US 20050089977 A1 20050428

APPLICATION INFO.: US 2004-990053 A1 20041115 (10)

RELATED APPLN. INFO.: Division of Ser. No. US 2001-870821, filed on 1 Jun 2001, PENDING

NUMBER DATE

PRIORITY INFORMATION: JP 2000-166578 20000602

JP 2000-166579 20000602

JP 2000-206130 20000707

DOCUMENT TYPE: Utility

FILE SEGMENT: APPLICATION

LEGAL REPRESENTATIVE: FINNEGAN, HENDERSON, FARABOW, GARRETT & DUNNER, LLP,

901 NEW YORK AVENUE, NW, WASHINGTON, DC, 20001-4413, US

NUMBER OF CLAIMS: 19

EXEMPLARY CLAIM: 1-10

NUMBER OF DRAWINGS: 4 Drawing Page(s)

LINE COUNT: 1853

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

AB The present invention relates to an enzyme reaction method which comprises performing an enzyme reaction, using an immobilized enzyme having a water content of 10% by weight or more as an enzyme and using an organic solvent substantially immiscible with water as a reaction solvent, under such conditions that a liquid phase forms a homogeneous system without phase separation although it is saturated with water or an aqueous buffer; a method for performing an enzyme reaction using an aldehyde compound as a substrate, which comprises removing a carboxylic acid compound contained in an aldehyde compound by subjecting the aldehyde compound to an alkaline treatment before starting the enzyme reaction; a method for performing an enzyme reaction using an aldehyde compound as a substrate, which comprises reducing a carboxylic acid compound content in the aldehyde compound to 0.1 wt % or less by subjecting the aldehyde compound to an alkaline treatment before starting the enzyme reaction; a method for enzymatically producing an optically active cyanohydrin from a carbonyl compound and prussic acid containing an acidic substance as a stabilizer, which comprises subjecting the prussic acid to a treatment for reducing inhibitory effect of the stabilizer on an enzyme, and performing an enzyme reaction to synthesize the optically active cyanohydrin using the treated prussic acid; a method for enzymatically producing an optically active cyanohydrin, which comprises dissolving prussic acid in an organic solvent substantially immiscible with water to give an organic solution of prussic acid, adding a buffer to this solution in a saturation amount or more, mixing, collecting the organic phase, and performing an enzyme reaction to synthesize the optically active cyanohydrin using the organic phase as prussic acid; as well as a method for enzymatically producing an optically active cyanohydrin, which comprises performing distillation of a reaction solution after completion of an enzyme reaction to separate and collect unreacted prussic acid and organic solvent therefrom, and repeatedly using the collected prussic acid and organic solvent at least once.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L7 ANSWER 6 OF 13 WPIDS COPYRIGHT 2008 THOMSON REUTERS on STN

ACCESSION NUMBER: 2005-725510 [74] WPIDS

DOC. NO. CPI: C2005-220821 [74]

TITLE: Novel ***modified*** S- ***hydroxynitrile***
lyase having improved heat tolerance, and
obtained by ***modifying*** amino acid in helix D3',
helix A and beta-sheet 2 domains in wild-type S-
hydroxynitrile ***lyase***, useful for
producing ***cyanohydrin***

DERWENT CLASS: B04; B05; D16

INVENTOR: HARAYAMA S; ICHIGE E; SEMBA H; SHIJKU T; HARAYAMA S I T;

SENBA T; SHIJKU T I T; YOSOKU T

PATENT ASSIGNEE: (JAPCO) NIPPON SEIKUBAI CO LTD; (DOKU-N) DOKURITSU

GYOSEI HOJIN SEIHIN HYOKA GIJU; (HARA-I) HARAYAMA S;

(ICHI-I) ICHIGE E; (SEMB-I) SEMBA H; (SHI-I) SHIJKU T

COUNTRY COUNT: 108

PATENT INFO ABBR.:

PATENT NO KIND DATE WEEK LA PG MAIN IPC

WO 2005095602 A1 20051013 (200574)* EN 120[21]
JP 2005312431 A 20051110 (200574) JA 32
EP 1730274 A1 20061213 (200701) EN
US 20080124784 A1 20080529 (200838) EN

APPLICATION DETAILS:

PATENT NO	KIND	APPLICATION	DATE
WO 2005095602 A1		WO 2005-JP6730	20050330
JP 2005312431 A		JP 2005-21721	20050128
EP 1730274 A1		EP 2005-728634	20050330
EP 1730274 A1		WO 2005-JP6730	20050330
US 20080124784 A1		WO 2005-JP6730	20050330
US 20080124784 A1		US 2006-594732	20060929

FILING DETAILS:

PATENT NO	KIND	PATENT NO
EP 1730274	A1 Based on	WO 2005095602 A

PRIORITY APPLN. INFO: JP 2004-105642 20040331

AN 2005-725510 [74] WPIDS

AB WO 2005095602 A1 UPAB: 20060125

NOVELTY - Modified S-hydroxynitrile lyase (SHNL) (I) obtained by modifying one or more amino acid in the helix D3', helix A, and beta-sheet 2 domains in the amino acid sequence of wild-type SHNL, and having any one of 13 fully defined 258 amino acid (SEQ ID No. 2, 6, 8, 16, 20, 22, 26, 28, 32, 36, 40, 42 or 44) sequences given in the specification, is new.

DETAILED DESCRIPTION - Modified S-hydroxynitrile lyase (SHNL) (I) that is obtained by modifying one or more amino acid in the helix D3', helix A, and beta-sheet 2 domains in the amino acid sequence of wild-type SHNL, comprises:

(a) one or more amino acid substitution in the amino acid sequence having a fully defined 258 amino acid (SEQ ID No. 2) sequence given in the specification, where the substitution is chosen from substitution from lysine to aspartic acid, glutamic acid or asparagine at position 21, substitution from glycine to aspartic acid or glutamic acid at position 165, substitution from valine to leucine at position 173, substitution from methionine to leucine at position 174, and substitution from threonine to aspartic acid, glutamic acid or serine at position 163, and
(b) has any one of 12 fully defined 258 amino acid (SEQ ID No. 6, 8, 16, 20, 22, 26, 28, 32, 36, 40, 42 or 44) sequences given in the specification.

INDEPENDENT CLAIMS are also included for:

(1) DNA (II) encoding the amino acid sequence of (I) or encoding SEQ ID No. 6, 8, 16, 20, 22, 26, 28, 32, 36, 40, 42 or 44;
(2) preparation of (I); and
(3) improving stability of (I), by modifying one or more amino acid in the helix D3', helix A, and beta-sheet 2 domains in the amino acid sequence of wild-type SHNL.

USE - (I) is useful for producing optically active cyanohydrin, which involves allowing (I) to react with a carbonyl compound and cyanide (claimed). (I) is useful for industrial production of optically active cyanohydrin. (I) is useful as an enzyme immobilized on an adequate inorganic carrier, and for synthesizing cyanohydrin.

ADVANTAGE - (I) has improved and excellent heat tolerance, stability and organic solvent tolerance. (I) can be purified easily and cost-effectively.

L7 ANSWER 7 OF 13 Elsevier BIOBASE COPYRIGHT 2008 Elsevier Science B.V. on STN

ACCESSION NUMBER: 2005166675 EMBASE <LOGIND:20081117>

TITLE: Inversion of stereoselectivity by applying mutants of the hydroxynitrile lyase from ***Manihot*** esculenta

AUTHOR: Buhler H.; Michlich B.; Effenberger F.
CORPORATE SOURCE: Dr. F. Effenberger, Institut für Organische Chemie, Universität Stuttgart, Pfaffenwaldring 55, 70569

Stuttgart, Germany.

E-mail: franz.effenberger@oc.uni-stuttgart.de

SOURCE: ChemBioChem, (2005), 6/4 (711-717), 31 reference(s)

CODEN: CBCHFX ISSN: 1439-4227

DOCUMENT TYPE: Journal; Article

COUNTRY: Germany, Federal Republic of

LANGUAGE: English

SUMMARY LANGUAGE: English

AB The influence of Trp128-^{***substituted***} mutants of the ^{***hydroxynitrile***} ^{***lyase***} from ^{***Manihot***} esculenta (MeHNL) on the stereoselectivity of MeHNL-catalyzed HCN additions to aldehydes with stereogenic centers, which yield the corresponding cyanohydrins, is described. In rac-2-phenylpropionaldehyde (rac-1) reactions, wild-type (wtMeHNL) and all MeHNL Trp128 mutants are highly (S)-selective toward the (R) enantiomer of rac-1; this results exclusively in (2S,3R)-^{***cyanohydrin***} ((2S,3R)-2) with >=96% de. The (S) enantiomer of rac-1, however, only reacts (S)-selectively with wtMeHNL to give (2S,3S)-2 with 80% de, whereas with Trp128 mutants, (R) selectivity increases with decreasing size of the amino acids exchanged. The MeHNL-W128A-^{***mutant***} is exclusively (R)-selective, resulting in (2R,3S)-2 with 86% de. The reaction behavior of rac-phenylbutyraldehyde (rac-5) is comparable with rac-1, which also inverts the stereoselectivity from (S) to (R) when the enzyme is exchanged from wtMeHNL to the W128A-^{***mutant***}. Stereogenic centers not adjacent to the aldehyde group, as in 7 and 9, do not influence the stereoselectivity of MeHNL catalysis, and (S) selectivity is observed in all cases. Stereoselectivity and inversion of stereoselectivity of MeHNL Trp128 ^{***mutant***}-catalyzed ^{***cyanohydrin***} formation can be explained and rationalized with crystal-structure-based molecular modeling. .COPYRGTF. 2005 Wiley-VCH Verlag GmbH & Co. KGaA.

L7 ANSWER 8 OF 13 Elsevier BIOBASE COPYRIGHT 2008 Elsevier Science B.V. on

STN DUPLICATE

ACCESSION NUMBER: 2003077165 ESBIOBASE <<LOGINID::20081117>>

TITLE: Substrate specificity of mutants of the hydroxynitrile

lyase from ^{***Manihot***} esculenta

AUTHOR: Buhler H.; Effenberger F.; Forster S.; Roos J.; Wajant

H.

CORPORATE SOURCE: Dr. F. Effenberger, Institut für Organische Chemie, Universität Stuttgart, Pfaffenwaldring 55, 70569

Stuttgart, Germany.

E-mail: franz.effenberger@po.uni-stuttgart.de

SOURCE: ChemBioChem, (03 MAR 2005), 4/2-3 (211-216), 30

reference(s)

CODEN: CBCHFX ISSN: 1439-4227

DOCUMENT TYPE: Journal; Article

COUNTRY: Germany, Federal Republic of

LANGUAGE: English

SUMMARY LANGUAGE: English

AB Several tryptophan 128-^{***substituted***} mutants of the ^{***hydroxynitrile***} ^{***lyase***} from ^{***Manihot***} esculenta (MeHNL) are constructed and applied in the MeHNL-catalyzed addition of HCN to various aromatic and aliphatic aldehydes as well as to methyl and ethyl ketones to yield the corresponding cyanohydrins. The mutants (especially MeHNL-W128A) are in most cases superior to the wild-type (wt) enzyme when diisopropyl ether is used as the solvent. ^{***Substitution***} of tryptophan 128 by an alanine residue enlarges the entrance channel to the active site of MeHNL and thus facilitates access of sterically demanding substrates to the active site, as clearly demonstrated for aromatic aldehydes, especially 3-phenoxybenzaldehyde. These experimental results are in accordance with the X-ray crystal structure of MeHNL-W128A. Aliphatic aldehydes, surprisingly, do not demonstrate this reactivity dependence of mutants on substrate bulkiness. Comparative reactions of 3-phenoxybenzaldehyde with wtMeHNL and MeHNL-W128A in both aqueous citrate buffer and a two-phase system of water/methyl tert-butyl ether again reveal the superiority of the ^{***mutant***} enzyme: 3-phenoxybenzaldehyde was converted quantitatively into a ^{***cyanohydrin***} nearly independently of the amount of enzyme present, with a space-time yield of 57 g L.sup.-sup.1 h.sup.-sup.1.

L7 ANSWER 9 OF 13 USPATFULL on STN
ACCESSION NUMBER: 2002:12267 USPATFULL <<LOGINID::20081117>>
TITLE: Enzyme reaction method and a method for enzymatically
producing an optically active cyanohydrin
INVENTOR(S): Semba, Hisashi, Ibaraki, Japan
Dobushi, Yukio, Ibaraki, JAPAN
PATENT ASSIGNEE(S): Nippon Shokubai Co., Ltd. (non-U.S. corporation)

NUMBER	KIND	DATE

PATENT INFORMATION:	US 20020006646	A1 20020117
US 7078225	B2 20060718	
APPLICATION INFO:	US 2001-870821	A1 20010601 (9)

NUMBER	DATE

PRIORITY INFORMATION:	JP 2000-166578 20000602
JP 2000-166579	20000602
JP 2000-206130	20000707
DOCUMENT TYPE:	Utility
FILE SEGMENT:	APPLICATION
LEGAL REPRESENTATIVE:	Finnegan, Henderson, Farabow, Garrett & Dunner, L.L.P., 1300 I Street, N.W., Washington, DC, 20005-3315
NUMBER OF CLAIMS:	21
EXEMPLARY CLAIM:	1
NUMBER OF DRAWINGS:	4 Drawing Page(s)
LINE COUNT:	1898
CAS INDEXING IS AVAILABLE FOR THIS PATENT.	

AB The present invention relates to an enzyme reaction method which comprises performing an enzyme reaction, using an immobilized enzyme having a water content of 10% by weight or more as an enzyme and using an organic solvent substantially immiscible with water as a reaction solvent, under such conditions that a liquid phase forms a homogeneous system without phase separation although it is saturated with water or an aqueous buffer; a method for performing an enzyme reaction using an aldehyde compound as a substrate, which comprises removing a carboxylic acid compound contained in an aldehyde compound by subjecting the aldehyde compound to an alkaline treatment before starting the enzyme reaction; a method for performing an enzyme reaction using an aldehyde compound as a substrate, which comprises reducing a carboxylic acid compound content in the aldehyde compound to 0.1 wt % or less by subjecting the aldehyde compound to an alkaline treatment before starting the enzyme reaction; a method for enzymatically producing an optically active cyanohydrin from a carbonyl compound and prussic acid containing an acidic substance as a stabilizer, which comprises subjecting the prussic acid to a treatment for reducing inhibitory effect of the stabilizer on an enzyme, and performing an enzyme reaction to synthesize the optically active cyanohydrin using the treated prussic acid; a method for enzymatically producing an optically active cyanohydrin, which comprises dissolving prussic acid in an organic solvent substantially immiscible with water to give an organic solution of prussic acid, adding a buffer to this solution in a saturation amount or more, mixing, collecting the organic phase, and performing an enzyme reaction to synthesize the optically active cyanohydrin using the organic phase as prussic acid; as well as a method for enzymatically producing an optically active cyanohydrin, which comprises performing distillation of a reaction solution after completion of an enzyme reaction to separate and collect unreacted prussic acid and organic solvent therefrom, and repeatedly using the collected prussic acid and organic solvent at least once.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L7 ANSWER 10 OF 13 Elsevier BIOBASE, COPYRIGHT 2008 Elsevier Science B.V. on STN DUPLICATE
ACCESSION NUMBER: 2002007378 ESBIOBASE <<LOGINID::20081117>>
TITLE: Structure determinants of substrate specificity of
hydroxynitrile lyase from ***Manihot*** esculenta
AUTHOR: Lauble H.; Michlich B.; Forster S.; Kobler C.; Wajant
H.; Effenberger F.

CORPORATE SOURCE: H. Lauble, Institut für Organische Chemie, Universität
Stuttgart, Pfaffenwaldring 55, D-70569 Stuttgart,
Germany.
E-mail: Peter.Lauble@t-online.de

SOURCE: Protein Science, (2002), 11/1 (65-71), 20 reference(s)
CODEN: PRClEI ISSN: 0961-8368

DOCUMENT TYPE: Journal; Article

COUNTRY: United States

LANGUAGE: English

SUMMARY LANGUAGE: English

AB Tryptophan 128 of ***hydroxynitrile*** ***lyase*** of
Manihot esculenta (MeHNL) covers a significant part of a
hydrophobic channel that gives access to the active site of the enzyme.
This residue was therefore ***substituted*** in the ***mutant***
MeHNL-W128A by alanine to study its importance for the substrate
specificity of the enzyme. Wild-type MeHNL and MeHNL-W128A showed
comparable activity on the natural substrate acetone ***cyanohydrin***
(53 and 40 U/mg, respectively). However, the specific activities of
MeHNL-W128A for the unnatural substrates mandelonitrile and
4-hydroxymandelonitrile are increased 9-fold and approx.450-fold,
respectively, compared with the wild-type MeHNL. The crystal structure of
the MeHNL-W128A substrate-free form at 2.1-ÅNG. resolution indicates
that the W128A ***substitution*** has significantly enlarged the
active-site channel entrance, and thereby explains the observed changes
in substrate specificity for bulky substrates. Surprisingly, the
MeHNL-W128A-4-hydroxybenzaldehyde complex structure at 2.1-ÅNG.
resolution shows the presence of two hydroxybenzaldehyde molecules in a
sandwich type arrangement in the active site with an additional hydrogen
bridge to the reacting center.

L7 ANSWER 11 OF 13 CAPLUS COPYRIGHT 2008 ACS on STN DUPLICATE 4

ACCESSION NUMBER: 2001:454037 CAPLUS <<LOGINID::20081117>>

DOCUMENT NUMBER: 135:164024

TITLE: Mechanistic aspects of cyanogenesis from active-site
mutant Ser80Ala of ***hydroxynitrile***
lyase from ***Manihot*** esculenta in
complex with acetone ***cyanohydrin***

AUTHOR(S): Lauble, Hanspeter; Miehlisch, Burkhard; Forster,
Siegfried; Wajant, Harald; Effenberger, Franz

CORPORATE SOURCE: Institut für Organische Chemie der Universität
Stuttgart, Stuttgart, D-70569, Germany

SOURCE: Protein Science (2001), 10(5), 1015-1022

CODEN: PRClEI; ISSN: 0961-8368

PUBLISHER: Cold Spring Harbor Laboratory Press

DOCUMENT TYPE: Journal

LANGUAGE: English

AB The structure and function of hydroxynitrile lyase from ***Manihot***
esculenta (MeHNL) were analyzed by x-ray crystallog. and site-directed
mutagenesis. The crystal structure of the MeHNL-S80A mutant was refined to
an R-factor of 18.0% against diffraction data to 2.1-ÅNG. resoln. The
3-dimensional structure of the MeHNL-S80A-acetone cyanohydrin complex was
dett. at 2.2-ÅNG. resoln. and refined to an R-factor of 18.7%. To
elucidate the role of Thr-11 and Cys-81 in the catalytic mechanism,
mutants T11A and C81A were constructed and kinetic values were detd. The
Km for mutant C81A was indistinguishable from that of the wild-type
enzyme, whereas that for mutant T11A could not be detd. because of its
poor enzymic activity. From these results, the roles of Cys-81 and Thr-11
in the enzyme were discussed. Combined with structural data, the results
supported a mechanism for cyanogenesis in which His-236 as a general base
abstrs. a proton from Ser-80, thereby allowing proton transfer from the OH
group of acetone cyanohydrin to Ser-80. The His-236 imidazolium cation
then facilitates the leaving of the nitrile group by proton donation.

REFERENCE COUNT: 23 THERE ARE 23 CITED REFERENCES AVAILABLE FOR THIS
RECORD. ALL CITATIONS AVAILABLE IN THE REFORMAT

L7 ANSWER 12 OF 13 CAPLUS COPYRIGHT 2008 ACS on STN

ACCESSION NUMBER: 2000:189013 CAPLUS <<LOGINID::20081117>>

Correction of: 2000:12726

DOCUMENT NUMBER: 132:191223

Correction of: 132:60999

TITLE: (S)-hydroxynitrile lyases with improved substrate binding arising from modifications affecting the substrate binding site and their uses

INVENTOR(S): Effenberger, Franz; Lauble, Peter; Buhler, Holger; Wajant, Harald; Forster, Siegfried; Schwab, Helmut; Kratky, Christoph; Wagner, Ulrike; Steiner, Ernst

PATENT ASSIGNEE(S): Dsm Fine Chemicals Austria GmbH, Austria

SOURCE: Eur. Pat. Appl., 6 pp.
CODEN: EPXXDW

DOCUMENT TYPE: Patent

LANGUAGE: German

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
EP 969095	A2	20000105	EP 1999-111574	19990615
EP 969095	A3	20020828		
EP 969095	B1	20040929		
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO				
AT 9801159	A	20000715	AT 1998-1159	19980702
AT 407397	B	20010226		
AT 278022	T	20041015	AT 1999-111574	19990615
ES 2227935	T3	20050401	ES 1999-111574	19990615
CA 2277594	A1	20000102	CA 1999-2277594	19990630
JP 2000125886	A	20000509	JP 1999-188094	19990701
US 6319697	B1	20011120	US 1999-345773	19990701
PRIORITY APPLN. INFO.: AT 1998-1159 A 19980702 EP 1999-111574 A 19990615				

AB Plant (S)-hydroxynitrile lyases that bind substrates more efficiently have modifications in the hydrophobic channel leading to the substrate binding site. The channel is modified by replacing some of the bulkier hydrophobic amino acids that partially block the channel with less bulky ones. Specifically, a tryptophan in the channel (position 128 in the Hevea brasiliensis and ***Manihot*** esculenta enzyme) may be substituted by alanine, glycine, valine, or even phenylalanine. The substituted M. esculenta converted 3-phenoxybenzaldehyde to (S)-3-phenoxybenzaldehyde cyanohydrin with an e.e. of 97% compared to 97.03% for the wild type enzyme. Yield of the product was 88.5% after 2h using the wild-type enzyme and 97% after 1.5 h using the substituted enzyme.

L7 ANSWER 13 OF 13 SCISEARCH COPYRIGHT (c) 2008 The Thomson Corporation on STN DUPLICATE 5

ACCESSION NUMBER: 1996:748457 SCISEARCH <LOGINID::20081117>>

THE GENUINE ARTICLE: VN180

TITLE: Identification of potential active-site residues in the hydroxynitrile Lyase from ***Manihot*** esculenta by site-directed mutagenesis

AUTHOR: Wajant H (Reprint); Pfizenmaier K

CORPORATE SOURCE: UNIV STUTTGART, INST CELL BIOL & IMMUNOL, ALLMANDRING 31, D-70569 STUTTGART, GERMANY (Reprint)

COUNTRY OF AUTHOR: GERMANY

SOURCE: JOURNAL OF BIOLOGICAL CHEMISTRY, (18 OCT 1996) Vol. 271, No. 42, pp. 25830-25834.
ISSN: 0021-9258.

PUBLISHER: AMER SOC BIOCHEMISTRY MOLECULAR BIOLOGY INC, 9650 ROCKVILLE PIKE, BETHESDA, MD 20814.

DOCUMENT TYPE: Article; Journal

FILE SEGMENT: LIFE

LANGUAGE: English

REFERENCE COUNT: 25

ENTRY DATE: Entered STN: 1996

Last Updated on STN: 1996

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

AB The ***hydroxynitrile*** ***lyase*** from ***cassava*** (***Manihot*** esculenta Crantz) (EC 4.1.2.37) catalyzes the decomposition of the achiral alpha-hydroxynitrile acetone ***cyanohydrin*** into HCN and acetone during cyanogenesis of damaged

plants. This enzyme can also be used for stereoselective synthesis of a wide array of (S)-cyanohydrins by addition of HCN to aldehydes or ketones. Optically active cyanohydrins are interesting intermediates for the synthesis of alpha-hydroxy acids, alpha-hydroxy ketones, or beta-ethanolamines, all of which are important building blocks in organic synthesis. Inhibition of ***hydroxynitrile*** ***lyase*** from *M. esculenta* (MeHNL) by serine- and histidine- ***modifying*** reagents suggests involvement of active site seryl and histidyl residues. Furthermore, serine 80 of MeHNL is part of the active site motif Gly-X-Ser-X-Gly/Ala, often considered as the hallmark of catalytic triads having independently evolved in four groups of enzymes: the alpha/beta hydrolase fold enzymes, subtilisins, the cysteine proteases, and the eukaryotic serine proteases. By site-directed mutagenesis, three residues critical for enzyme activity have been identified: serine 80, aspartic acid 208, and histidine 236. These residues may be directly involved in MeHNL-catalyzed decomposition of cyanohydrins, providing evidence for a catalytic triad in HNLs, too. The order of the catalytic triad residues in the primary sequence of MeHNL is nucleophile-histidine-acid, suggesting that MeHNL belongs to the alpha/beta hydrolase fold group of enzymes. In contrast to all other enzymes having a catalytic triad, HNLs catalyze no net hydrolytic reactions.

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SEA ((HYDROXYNITRILE (W)LYASE) OR HYDROXYNITRILASE)

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1   FILE ANTE
44  FILE BIOENG
159 FILE BIOSIS
100 FILE BIOTECHABS
100 FILE BIOTECHDS
59  FILE BIOTECHNO
56  FILE CABA
319 FILE CAPLUS
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84  FILE ESBIOBASE
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87  FILE MEDLINE
71  FILE PASCAL
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240 FILE SCISEARCH
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132 FILE USGENE
76  FILE USPATFULL
1   FILE USPATOLD

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18 FILE USPAT2
46 FILE WPIDS
1 FILE WPIFV
46 FILE WPIINDEX
1 FILE NLDB
L1 QUE ((HYDROXYNITRILE (W) LYASE) OR HYDROXYNITRILASE)

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ON 17 NOV 2008

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L4 44 S CYANOHYDRIN (S) L3
L5 23 S (CASSAVA OR MANIHOT) (S) L4
L6 29 S (CASSAVA OR MANIHOT) AND L4
L7 13 DUP REM L6 (16 DUPLICATES REMOVED)

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